

# Haplotypes and databases

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## ● WORKSHOP 1

X-chromosomal markers in forensic genetics  
Daniel Kling & Andreas Tillmar

## ● WORKSHOP 2

Acreditación en el campo de la Genética Forense y  
estrategias de validación de ensayos  
Manuel Crespillo Márquez, Rosalía Izquierdo & Estel Enricig Cabanes

## ● WORKSHOP 3

La genética en la Identificación de víctimas a gran escala:  
comparación de perfiles y evaluación estadística con Familias  
Carlos Vullo & Lourdes Prieto

# Teachers

Daniel Kling, PhD



- Forensic Expert
- National Board of Forensic Medicine, Sweden
- Worked in the field for almost 15 years
- Developer of Familias, FamLink and FamLinkX
- Applied biostatistics, relationship inference, genetic genealogy

Andreas Tillmar, PhD



- Forensic geneticist & Associate professor
- National Board of Forensic Medicine, Sweden and Linköping University, Sweden
- Worked in the field for almost 20 years
- Technical leadership mixed with R&D
- Applied biostatistics, relationship inference, population genetics, genetic genealogy

## **Disclaimer!**

Points of view are those of the presenters and do not necessarily represent the official position or policies of the National Board of Forensic Medicine or ISFG. Certain commercial software, instruments, and materials are identified in order to specify experimental procedures as completely as possible. In no case does such identification imply a recommendation or endorsement, nor does it imply that any of the materials, instruments, or equipment identified are necessarily the best available for the purpose.

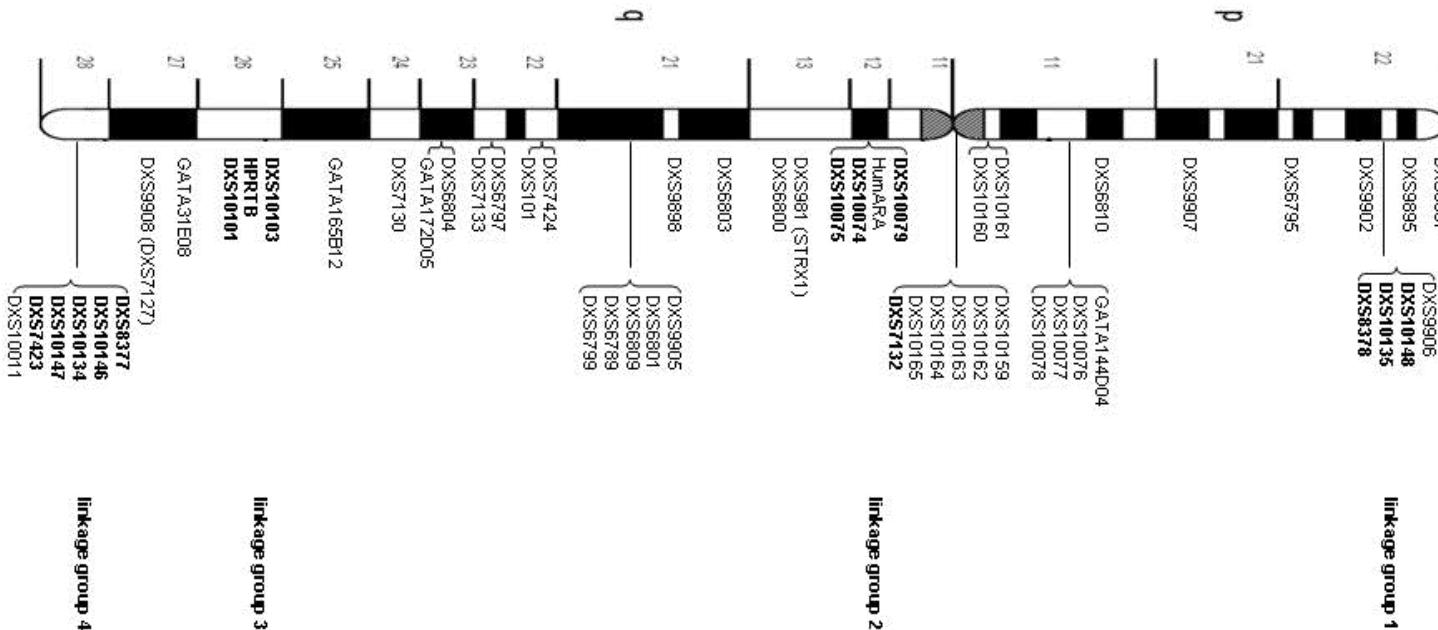
# Topics

- Markers
- Haplotypes
- Exploring population genetics of X-STRs

# Markers

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# Markers



# Markers

## Argus X12

Cytogenetic localisation	Marker	Linkage group	Physical localisation	Rutgers Map v.2
			[Mb]	[cM Kosambi] NCBI 36
			NCBI 36	
p 22.31	<a href="#">DXS10148</a>	X1	9.198	19.84
p 22.31	<a href="#">DXS10135</a>	X1	9.199	20.03
p 22.31	<a href="#">DXS8378</a>	X1	9.33	20.21
centromere	<a href="#">DXS7132</a>	X2	64.572	90.75
q 12	<a href="#">DXS10079</a>	X2	66.632	90.82
q 12	<a href="#">DXS10074</a>	X2	66.894	90.83
q 26.2	<a href="#">DXS10103</a>	X3	133.246	149.37
q 26.2	<a href="#">HPRTB</a>	X3	133.443	149.66
q 26.3	<a href="#">DXS10101</a>	X3	133.482	149.75
q 28	<a href="#">DXS10146</a>	X4	149.335	183.72
q 28	<a href="#">DXS10134</a>	X4	149.401	183.96
q 28	<a href="#">DXS7423</a>	X4	149.46	184.19

<https://pubmed.ncbi.nlm.nih.gov/24632058/>

## Decaplex

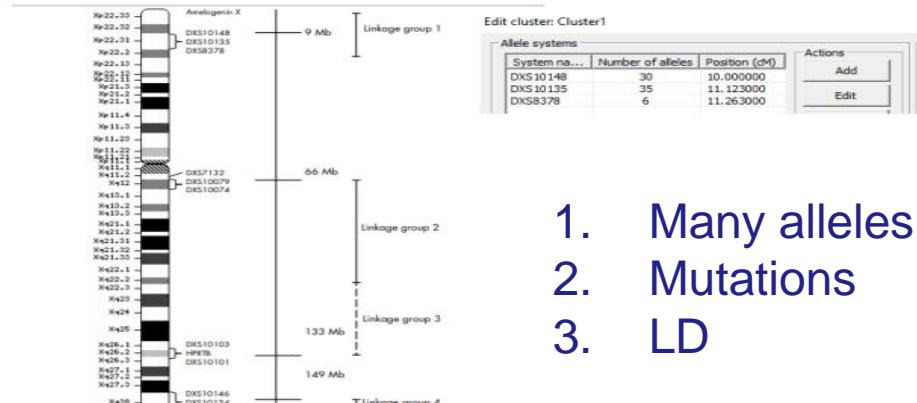
Cytogenetic localisation	Marker	Physical localisation	Rutgers Map v.2
		[Mb]	[cM Kosambi] NCBI 36
		NCBI 36	
p 22.31	<a href="#">DXS8378</a>	9.33	20.21
p 22.2	<a href="#">DXS9902</a>	15.234	32.32
centromere	<a href="#">DXS7132</a>	64.572	90.75
q 21.31	<a href="#">DXS9898</a>	87.682	101.29
q 21.33	<a href="#">DXS6809</a>	94.825	108.12
q 21.33	<a href="#">DXS6789</a>	95.336	108.47
q 22.3	<a href="#">DXS7133</a>	108.928	118.18
q 23	<a href="#">GATA172D05</a>	113.061	124.36
q 27.1	<a href="#">GATA31E08</a>	140.062	160.54
q 28	<a href="#">DXS7423</a>	149.46	184.19

<https://pubmed.ncbi.nlm.nih.gov/19082839/>

} <0.5cm

# Markers

- Different kits
  - STRs

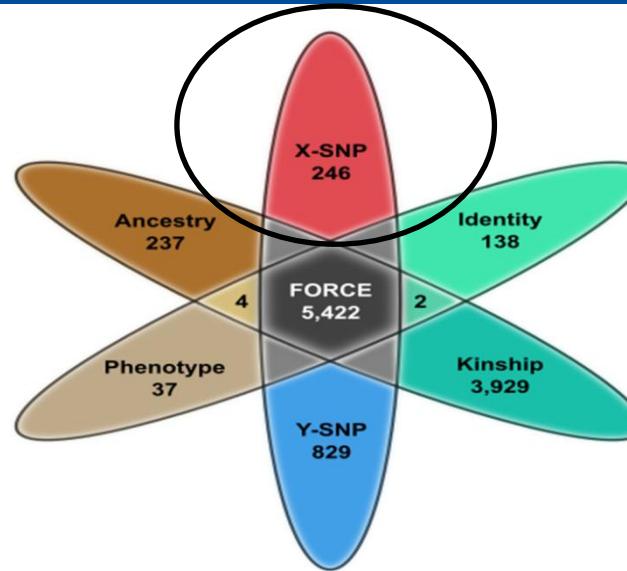


1. Many alleles
2. Mutations
3. LD

Figure 1. The ideogram of the X-chromosome describes the physical localization of the STR loci that can be analyzed using the Investigator Argus X-12 Q5 Kit. Distances from the p telomere are shown in Mb. ([www.ncbi.nlm.nih.gov/genome/guide/human](http://www.ncbi.nlm.nih.gov/genome/guide/human) as of 11/2014).

# Markers

1	Marker	Genetic position (cM)
2	rs4892897	7.582
3	rs1637781	7.805
4	rs5983084	8.5874
5	rs6642174	8.8134
6	rs6641753	9.1834
7	rs6641574	9.4251
8	rs2058865	9.7825
9	rs5962087	11.3138
10	rs5915796	12.2585
11	rs5916138	12.71
12	rs5915672	13.1805
13	rs6529997	13.9103
14	rs1637788	15.1505
15	rs4240138	15.6932
16	rs2108400	16.1319
17	rs5933710	17.0167
18	rs845444	17.6324
19	rs768568	17.9229
20	rs929217	18.1746



Open Access Article

The FORCE Panel: An All-in-One SNP Marker Set for Confirming Investigative Genetic Genealogy Leads and for General Forensic Applications

by <sup>1</sup> Andreas Tillmar <sup>1,2,\*</sup>, <sup>1</sup> Kimberly Sturk Andreaggi <sup>3,4,5</sup>, <sup>1</sup> Jennifer Daniels-Higginbotham <sup>3,4</sup>, <sup>2</sup> Jacqueline Tyler Thomas <sup>3,4</sup> and <sup>2</sup> Charla Marshall <sup>3,4,6,\*</sup>

<sup>1</sup> Department of Forensic Genetics and Forensic Toxicology, National Board of Forensic Medicine, SE-587 58 Linköping, Sweden

<sup>2</sup> Department of Biomedical and Clinical Sciences, Faculty of Medicine and Health Sciences, Linköping University, SE-582 25 Linköping, Sweden

<sup>3</sup> Armed Forces Medical Examiner System's Armed Forces DNA Identification Laboratory (AFMES-AFDIL), Dover Air Force Base, Dover, DE 19902, USA

<sup>4</sup> SNA International, LLC, Contractor Supporting the AFMES-AFDIL, Alexandria, VA 22314, USA

<sup>5</sup> Department of Immunology, Genetics and Pathology, Uppsala University, SE-751 08 Uppsala, Sweden

X-SNPs are chosen to be have no significant LD

# Haplotypes

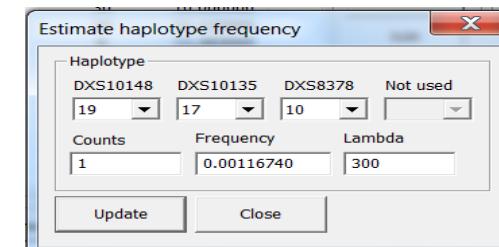
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# Haplotypes

- What is a haplotype? Sequence of alleles on the same chromosome
- What is special with the X-chromosome? Males only have one
- Phase uncertainty

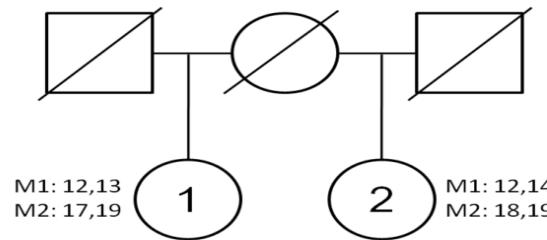
Marker	Child	Sibling
DXS10148	19,20	19 19
DXS10135	16,17	16 17
DXS8378	10,10	10 10

Haplotype



# Haplotypes

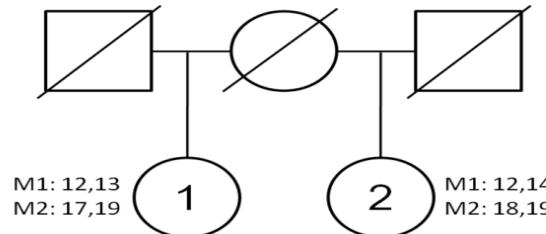
## Ambigious haplotypes



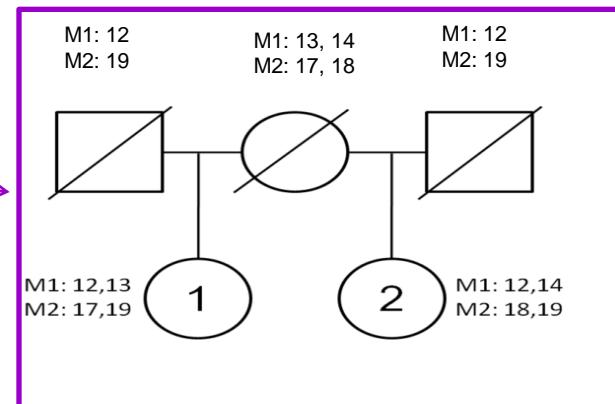
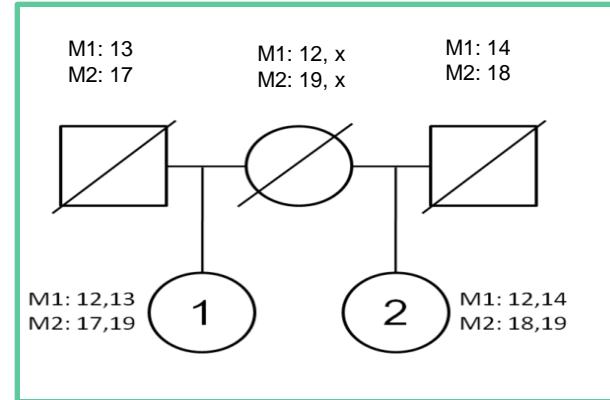
12\_19 can be from mother or  
from different fathers

# Haplotypes

## Ambigious haplotypes



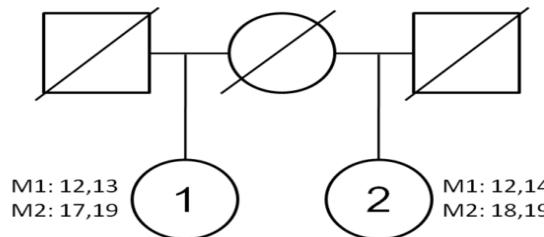
12\_19 can be from **mother** or  
from **different fathers**



Other possibilities?

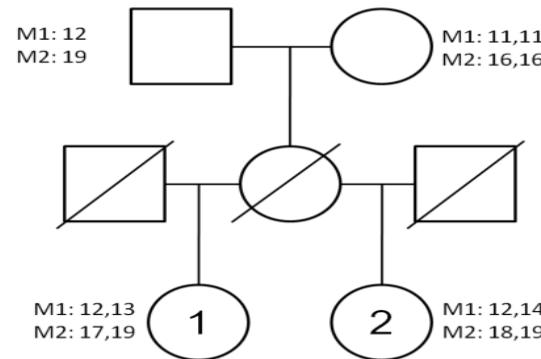
# Haplotypes

## Ambigious haplotypes



12\_19 can be from mother or  
from different fathers

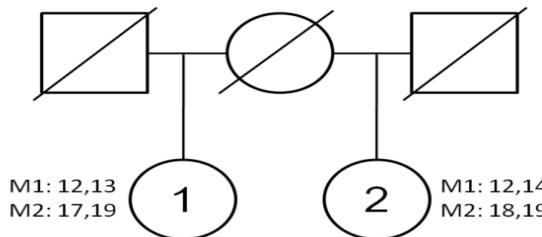
## Haplotypes "certain"



12\_19 is from the grandfather

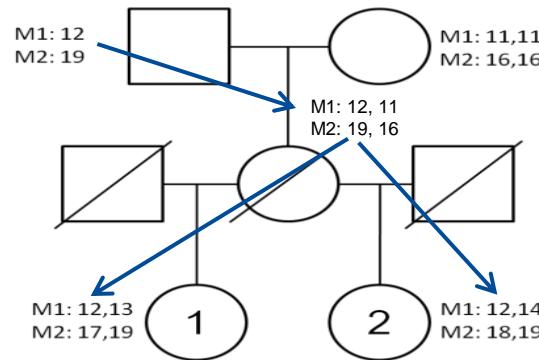
# Haplotypes

## Ambigious haplotypes



12\_19 can be from mother or from different fathers

## Haplotypes "certain"



12\_19 is from the grandfather

# Haplotypes

## Ambigious haplotypes

	DXS10103	DXS8378	DXS7132	DXS10134	DXS10074	DXS10101	DXS10135	DXS7423	DXS10146	DXS10079	HPRTB	DXS10148
Mother	19	10	13	37	16	28.2	18	14	28	20	12	24.1
	21	11	14	38	17	30.2	23	16	30	21	12	24.1
Son 1	19	10	13	37	16	28.2	18	14	28	20	12	24.1
Son 2	21	11	14	37	16	28.2	18	14	28	20	12	24.1

One crossover

# Haplotypes

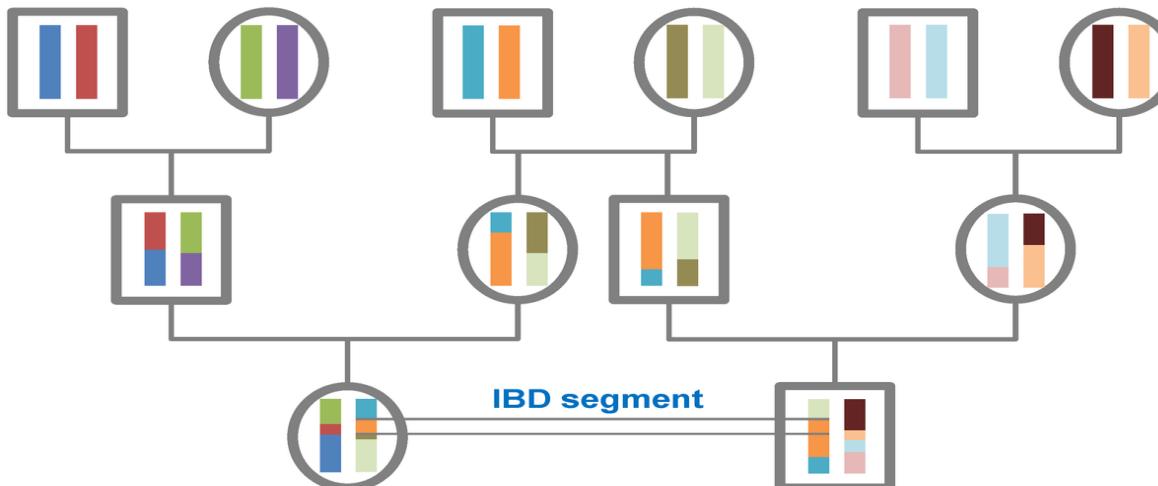
## Ambigious haplotypes

	DXS10103	DXS8378	DXS7132	DXS10134	DXS10074	DXS10101	DXS10135	DXS7423	DXS10146	DXS10079	HPRTB	DXS10148
Grandfather	19	10	13	37	17	28.2	23	16	30	21	12	24.1
Mother	19	10	13	37	16	28.2	23	16	30	21	12	24.1
	21	11	14	38	17	30.2	18	14	28	20	12	24.1
Son 1	19	10	13	37	16	28.2	18	14	28	20	12	24.1
Son 2	21	11	14	37	16	28.2	18	14	28	20	12	24.1

Three crossovers!

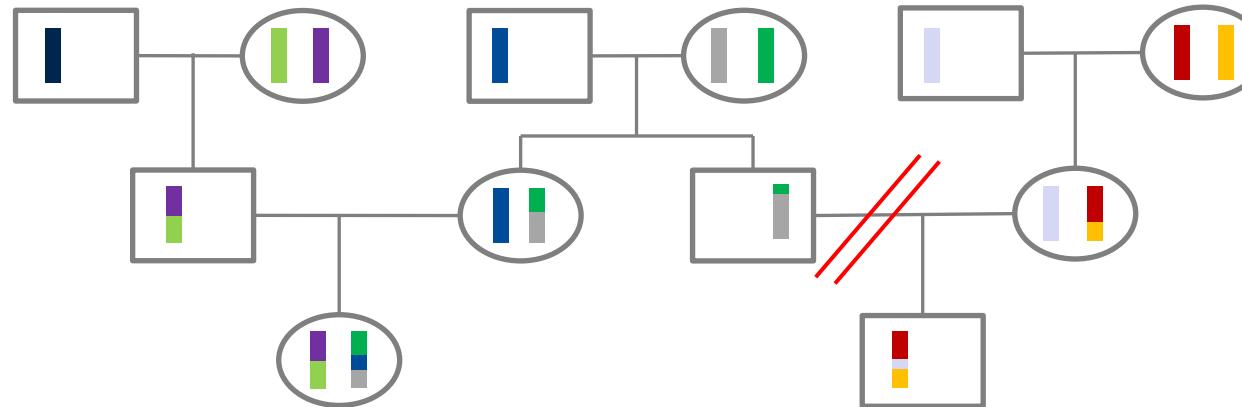
# Haplotypes

Tracing X-haplotypes through pedigrees



# Haplotypes

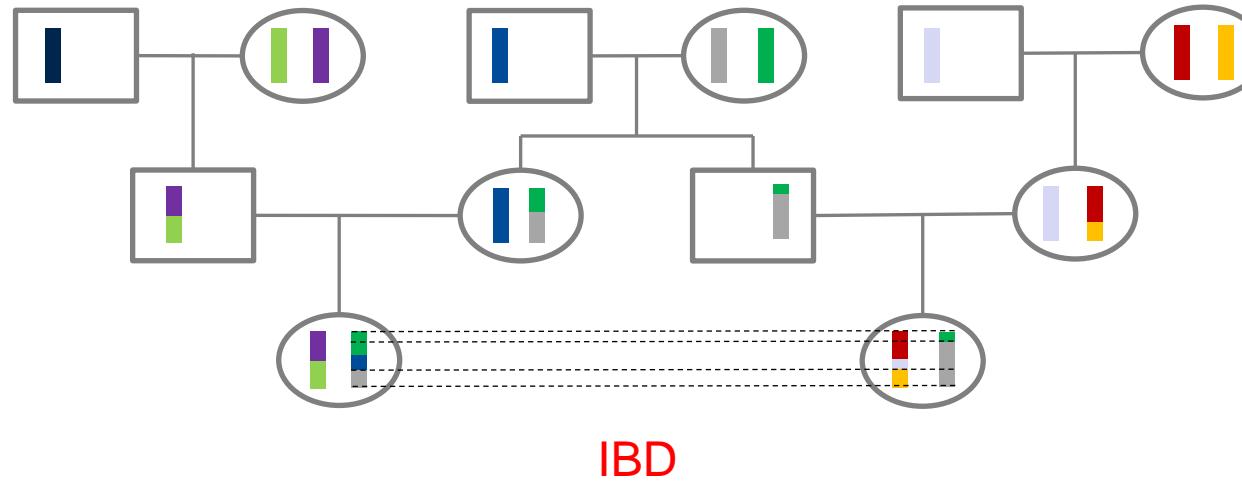
Tracing haplotypes through pedigrees



No IBD possible!

# Haplotypes

Tracing X-haplotypes through pedigrees



# Haplotypes

- For X-chromosomal databasing – males are preferred
- Exact inference of haplotypes

Marker	Male1	Male2	Male3	Male4	...	Male <i>n</i>
DXS10148	19	19	20	20	...	23
DXS10135	16	16	16	17	...	17
DXS8378	10	10	11	11	...	11

# Haplotypes



Forensic Science International: Genetics

Volume 15, March 2015, Pages 127-130



## Expanding X-chromosomal forensic haplotype frequencies database: Italian population data of four linkage groups

Carla Bini <sup>a</sup>  , Laura Natalia Riccardi <sup>a</sup>, Stefania Ceccardi <sup>a</sup>, Francesco Carano <sup>a</sup>, Stefania Sarno <sup>b</sup>, Donata Luiselli <sup>b</sup>, Susi Pelotti <sup>a</sup>

 [Show more](#)

<https://doi.org/10.1016/j.fsigen.2014.11.008>

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### Highlights

- X-chromosome STRs analysis has a potential usefulness in paternity testing.
- Haplotype frequencies in 200 unrelated Italian males have been estimated.
- Comparison with European populations showed some minor differences.
- Contribute to the statistical evaluation of the X-STRs polymorphism.

Bini et al., *Forensic Science International: Genetics*, Vol. 15, p127–130

LG I			
DXS8378	DXS10135	DXS10148	Number
8	29	27,1	1
9	22	24,1	1
10	19	23,1	1
10	19	24,1	2
10	19	25,1	2
10	19	26,1	2
10	19,1	18	1
10	20	24,1	1
10	20,1	25,1	1
10	20,1	26,1	1
10	20,1	27,1	1
10	20,1	28,1	1
10	21	20	1
10	21	23	1
10	21	26,1	2
10	21,1	18	1
10	21,1	26,1	2
10	22	18	1
10	22	21	1
10	22	24,1	2
10	22	26,1	2

LG II			
DXS7132	DXS10074	DXS10079	Number
10	17	19	1
11	8	20	1
11	17	20	1
11	19	21	1
12	7	19	2
12	8	18	2
12	8	20	1
12	8	21	1
12	9	21	1
12	16	18	1
12	16	20	1
12	17	18	2
12	18	16	1
12	18	18	1
12	18	19	3
12	18	22	1
12	19	19	1
12	20	20	1
13	7	20	2
13	7	21	1

LG III			
DXS10103	DXS10101	HPTB	Number
15	33	12	1
16	26	14	1
16	28,2	12	3
16	28,2	14	1
16	30	12	1
16	31	14	1
16	31,2	14	1
16	32	12	1
16	34	14	1
17	26,2	12	1
17	28	12	1
17	29	13	1
17	30	13	2
17	30	14	2
17	30	15	2
17	30,2	16	1
17	31	13	4
17	31	14	3
17	31	15	3
17	31,2	12	1

# Haplotypes

famlink.se/fx\_databases.html

## FamLinkX – A software for X-chromosomal markers

[Home](#) [Download](#) [Databases](#) [Getting started](#) [Manual](#) [Contact](#) [Help](#)

### Databases

The files below are prepared based on data given in the references. Please keep in mind that we have not performed quality control of the submitted data.

Send us a request on new databases or merging of several of the databases below.

All data for the Decaplex kit is obtained via Supplementary data in [Gusmão et al.](#).

**Frequency databases (haplotypes and allele frequencies)**

Databases may be found [here](#) or downloaded in the tables below. Size refers to the number of male haplotypes in the publication.

Please note that several populations can appear twice (different sources) and in continental databases as well.

### Argus X12

Population	Size	Database	Publication	Comment	Kit
Sweden	652	SWE	<a href="#">Tillmar A.</a>		Argus X12
Norway	631	NOR	<a href="#">Bergseth et al.</a>		Argus X12
Czech Republic	307	CZE	<a href="#">Zidkova et al.</a>		Argus X12
Germany	1037	GER	<a href="#">Edelmann et al.</a>		Argus X12
Greece	121	GRE	<a href="#">Tomas et al.</a>		Argus X12
Italian	200	ITA	<a href="#">Bini et al.</a>		Argus X12
Sardinia	316	SAR	<a href="#">Robino et al.</a>		Argus X12
Serbia	220	SER	<a href="#">Veselinovic et al.</a>		Argus X12

# Haplotypes

- Estimating haplotype frequencies
  - A long history of different methods
    - Expectation maximization (EM)
    - MCMC - PHASE
    - IMPUTE2, SHAPEIT2
- For X-chromosomal STR markers we consider a Dirichlet model

# Haplotypes

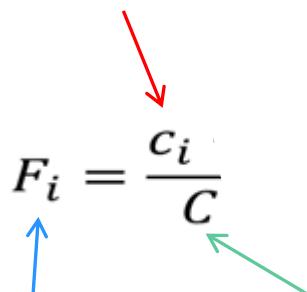
*The Lambda model*

$$F_i = \frac{c_i}{C}$$

Observations

Haplotype frequency

Size of database



# Haplotypes

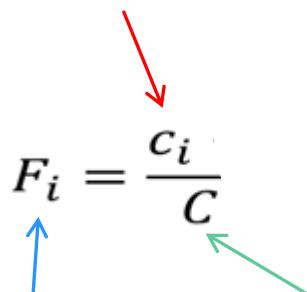
*The Lambda model*

$$F_i = \frac{c_i}{C}$$

Observations

Haplotype frequency

Size of database



Example

Haplotype 12\_20 is observed 10 times in a database of size 652

$$F_{12\_20} = 10/652$$

# Haplotypes

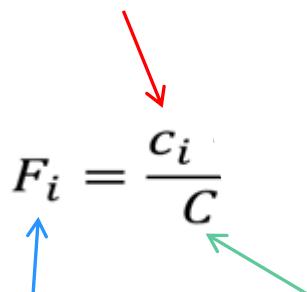
*The Lambda model*

$$F_i = \frac{c_i}{C}$$

Observations

Haplotype frequency

Size of database



Example

Haplotype 12\_20 is observed 0 times in a database of size 652

$$F_{12\_20} = 0/652$$

# Haplotypes

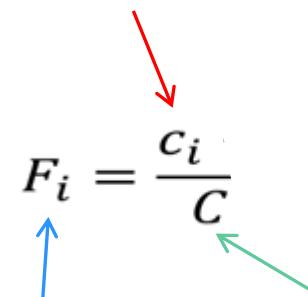
*The Lambda model*

$$F_i = \frac{c_i}{C}$$

Observations

Haplotype frequency

Size of database

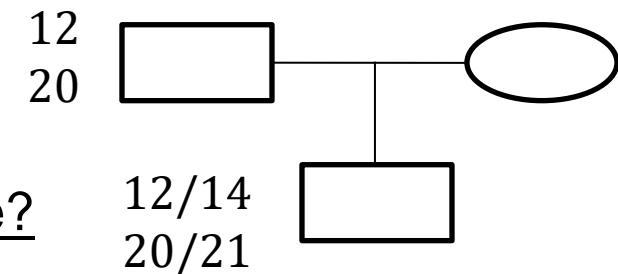


What about this case?

Example

Haplotype 12\_20 is observed 0 times in a database of size 652

$$F_{12\_20} = 0/652$$



# Haplotypes

## The Lambda model

$$F_i = \frac{c_i + \lambda\pi_i}{C + \lambda}$$

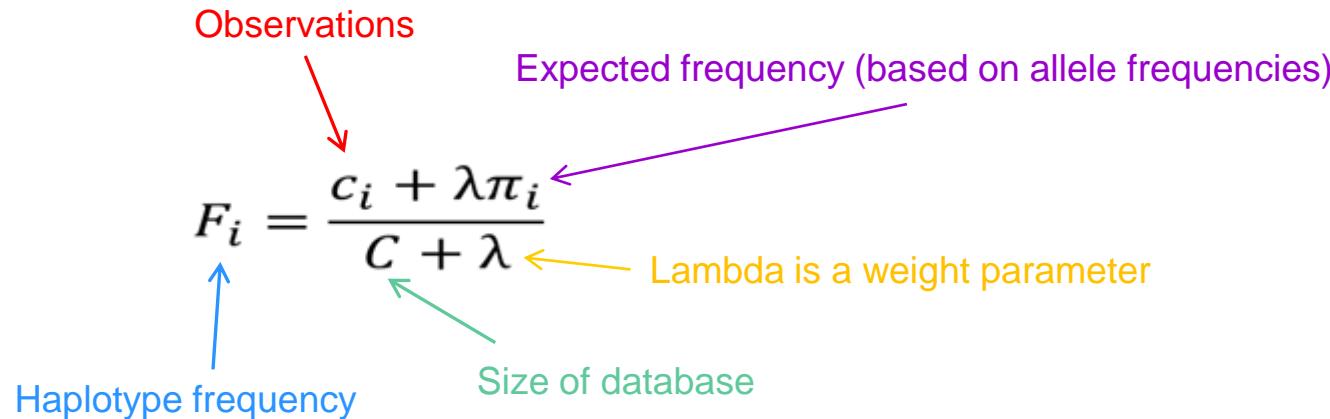
Observations

Expected frequency (based on allele frequencies)

Haplotype frequency

Size of database

Lambda is a weight parameter



The diagram illustrates the Lambda model equation. At the center is the formula  $F_i = \frac{c_i + \lambda\pi_i}{C + \lambda}$ . Five arrows point to different parts of the equation: a red arrow from 'Observations' to  $c_i$ ; a purple arrow from 'Expected frequency (based on allele frequencies)' to  $\lambda\pi_i$ ; a blue arrow from 'Haplotype frequency' to  $C$ ; a green arrow from 'Size of database' to  $\lambda$ ; and an orange arrow from 'Lambda is a weight parameter' to the  $\lambda$  term.

# Haplotypes

## The Lambda model

$$F_i = \frac{c_i + \lambda \pi_i}{C + \lambda}$$

Observations →

Expected frequency (based on allele frequencies) ←

Haplotype frequency ↑

Size of database ←

Lambda is a weight parameter

$F_{12\_20}=?$

What about this case?

12      20

12/14      20/21

```
graph LR; M12["12  
20"] --- C1214["12/14"]; M20["20"] --- C1214; C1214 --- Child[""];
```

## Example

Haplotype 12\_20 is observed 0 times in a database of size 652

# Haplotypes

## The Lambda model

$$F_i = \frac{c_i + \lambda \pi_i}{C + \lambda}$$

Observations → Expected frequency (based on allele frequencies)

↑ Haplotype frequency      ← Lambda is a weight parameter

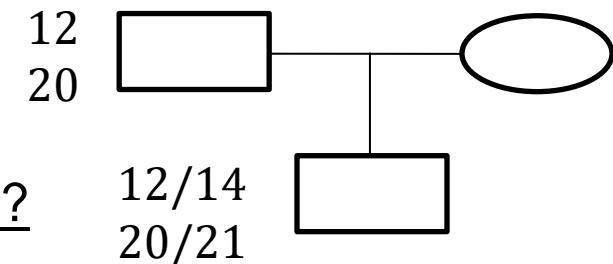
← Size of database

What about this case?

## Example

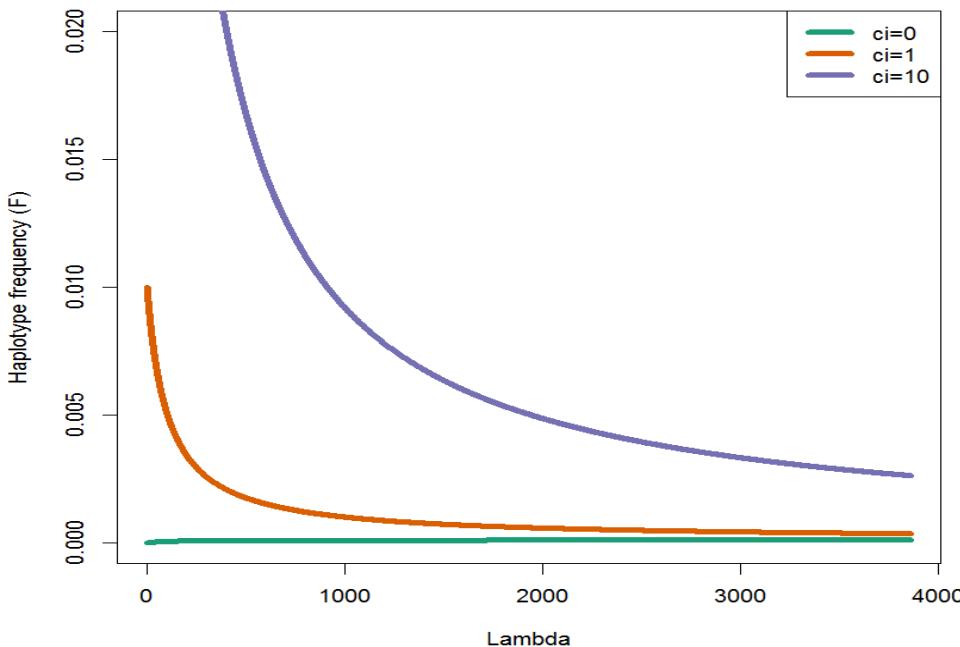
Haplotype 12\_20 is observed 0 times in a database of size 652. Lambda=100, p12=0.2, p20=0.3

$$F_{12\_20} = (0 + 100 * 0.2 * 0.3) / (652 + 100) = 6/752$$



# Haplotypes

$$F_i = \frac{c_i + \lambda\pi_i}{C + \lambda}$$



Marker	Haplotype
DXS10148	19
DXS10135	16
DXS8378	10

$c_i$	$F_i(\lambda=0)$	$F_i(\lambda=4000)$	$F_i(\lambda=\infty)$
0	0	0.0001	0.0001
1	1/100	0.00035	0.0001
10	10/100	0.0025	0.0001

C=652

# Difficulties

- In computations of likelihoods we,
  - ❑ Model population haplotype diversity – Estimate frequencies
  - ❑ Model haplotype transmission within a pedigree – Recombination
- Requires iteration over possible haplotype «states» - Phasing
- Mutations are highly relevant for STR markers (less so for SNPs)

More next week!

*Haplotypes are not needed for statistical calculation when using markers from Decaplex (or FORCE)!*

*Instead, allele frequencies are sufficient*

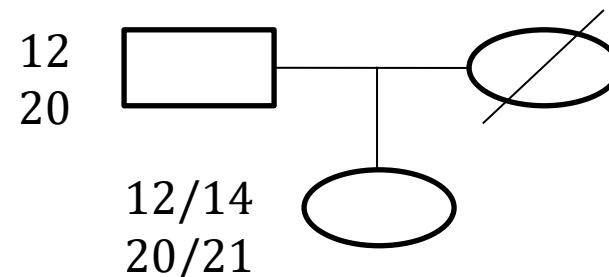
# FamLinkX – Demonstration 4

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# Demonstration

Marker	Position (cM)	Alleles	Frequencies
DXS10148	19.84	12, 14	0.1, 0.9
DXS10135	20.03	20, 21	0.1, 0.9

Haplotype	Observations	Freq
12 20	1	1/100
12 21	9	9/100
14 20	1	1/100
14 21	89	89/100



$$LR_{af} = 1/2p_{12} * 1/2p_{20} = 5 * 5 = \underline{25}$$

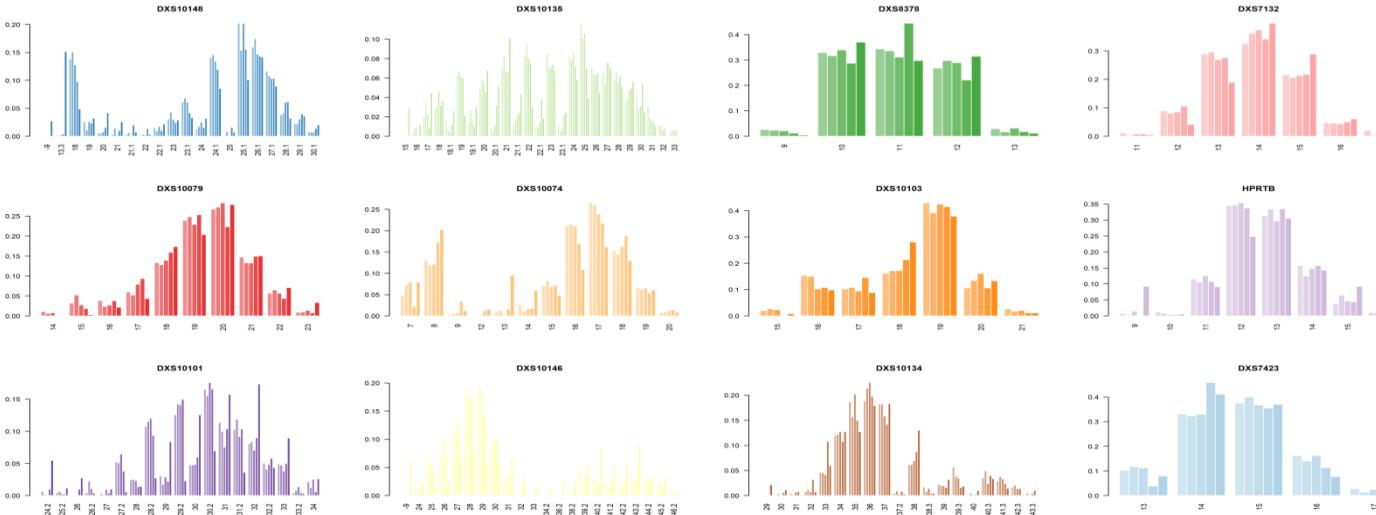
$$LR_{hf} = p_{14,21} / (2p_{12,20}p_{14,21} + 2p_{12,21}p_{14,20}) = 0.89 / 2(0.01 * 0.89 + 0.09 * 0.01) = \underline{45}$$

# Databases and populations

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# Population genetics

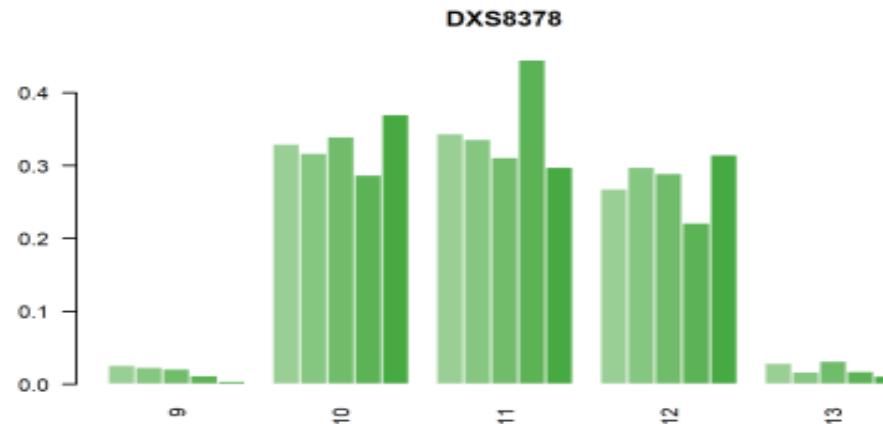
Allele frequencies for 5 different populations



Interpretation: Little information in allele frequencies alone?

# Population genetics

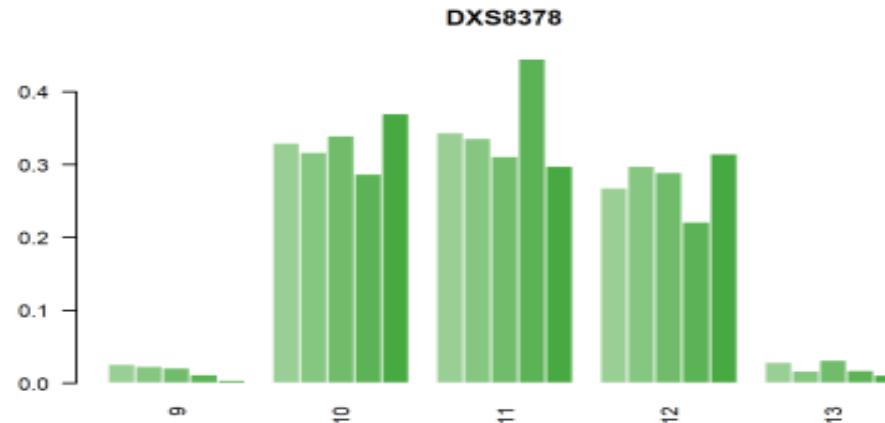
Allele frequencies for 5 different populations



Interpretation: Little information in allele frequencies alone?

# Population genetics

Allele frequencies for 5 different populations



Interpretation: Little information in allele frequencies alone?

# STRUCTURE

Each sample is assigned a mosaic of populations assuming K populations

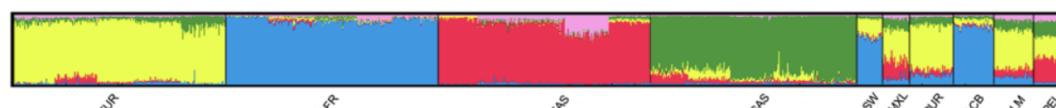
K = 3



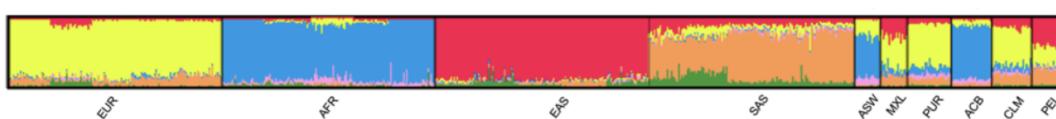
K = 4



K = 5

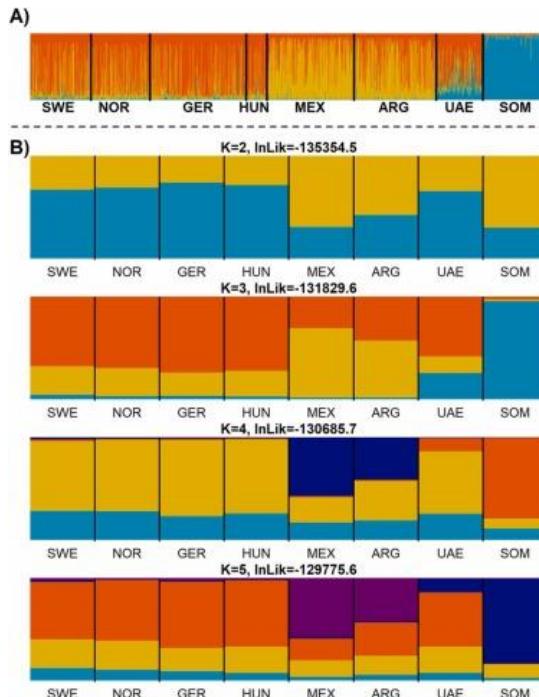


K = 6



E.g. yellow represents a European ancestry

# STRUCTURE



Interpretation: A vague trend where Somalia is represented by blue-ish. Too few markers to draw strong conclusions about population differences using this method.

  
**Forensic Science International: Genetics**  
 Volume 60, September 2022, 102745

Research paper

Extended population genetic analysis of 12 X-STRs – Exemplified using a Norwegian population sample

Erik F. Bergseth <sup>a,1</sup>, Andreas Tillmar <sup>b,c,1</sup>, P. Jørgen T. Hoddeland <sup>a</sup>, Daniel Kling <sup>a,b</sup>  

Show more 

+ Add to Mendeley  Cite

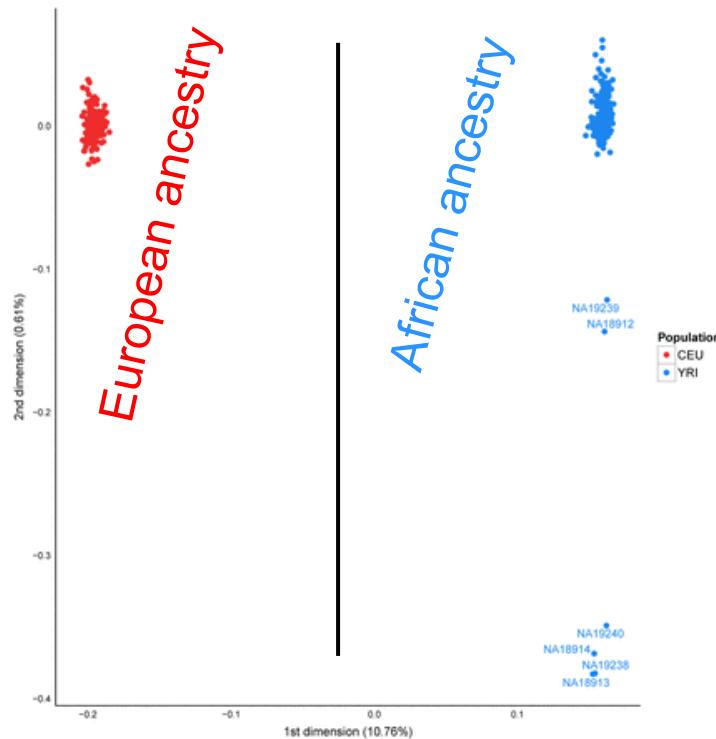
<https://doi.org/10.1016/j.fsigen.2022.102745> 

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**Highlights**

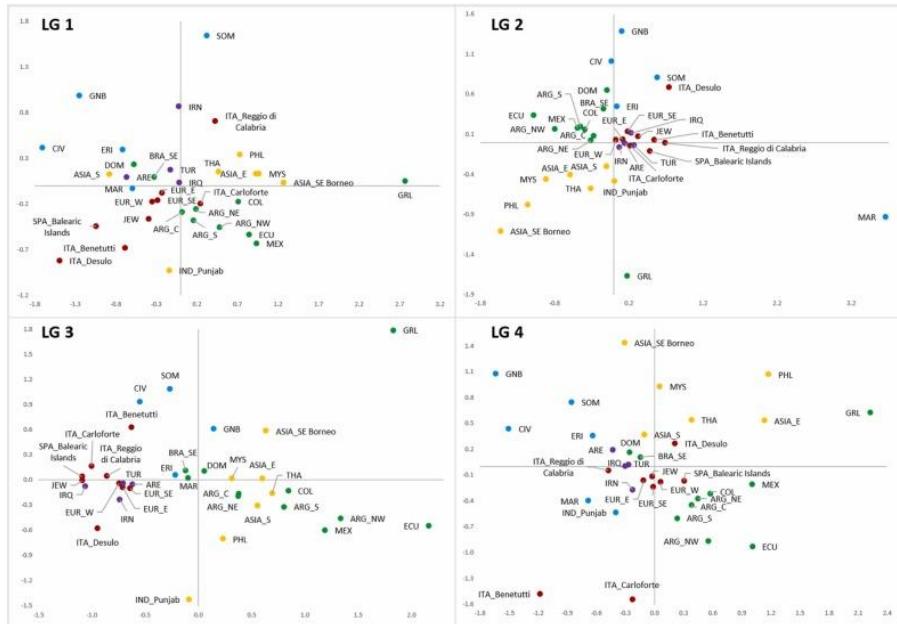
- Untraditional approaches to evaluating forensic markers.
- Exploring the LD structure of X-chromosomal haplotype data.
- Contrasting p-values with entropy to visualize LD.
- Contrasting the potential of X-chromosomal and autosomal STR markers.

# Differentiation



Separates multi-dimensional data into two Dimensions (MDS or PCA)

# Differentiation



*From Gusmao et al. 2025*

Interpretation: A vague separation with outliers

 ELSEVIER

Forensic Science International: Genetics  
Volume 76, March 2025, 103232



# X-chromosomal STRs: Metapopulations and mutation rates

L. Gusmão<sup>a,1</sup>, S. António-Sousa<sup>b,c,d,1</sup>, M. Faustino<sup>c,d</sup>, M.A. Abivórfiz<sup>d</sup>, D. Aguirre<sup>e</sup>, R. Alghofri<sup>f</sup>, C. Alves<sup>a</sup>, A. Amorim<sup>b,c</sup>, C. Arevalo<sup>b</sup>, L. Baldassari<sup>c</sup>, C. Barletto-Carroll<sup>c</sup>, G. Berardi<sup>b</sup>, C. Bobillo<sup>a</sup>, L. Bojar<sup>a</sup>, D.F. Brognoli<sup>b</sup>, A. Brehm<sup>c</sup>, J.J. Builes<sup>c</sup>, L. Carné<sup>b,c</sup>, E.F. Carvalho<sup>a</sup>, M. Carvalho<sup>a</sup>, N. Pinto<sup>a</sup>

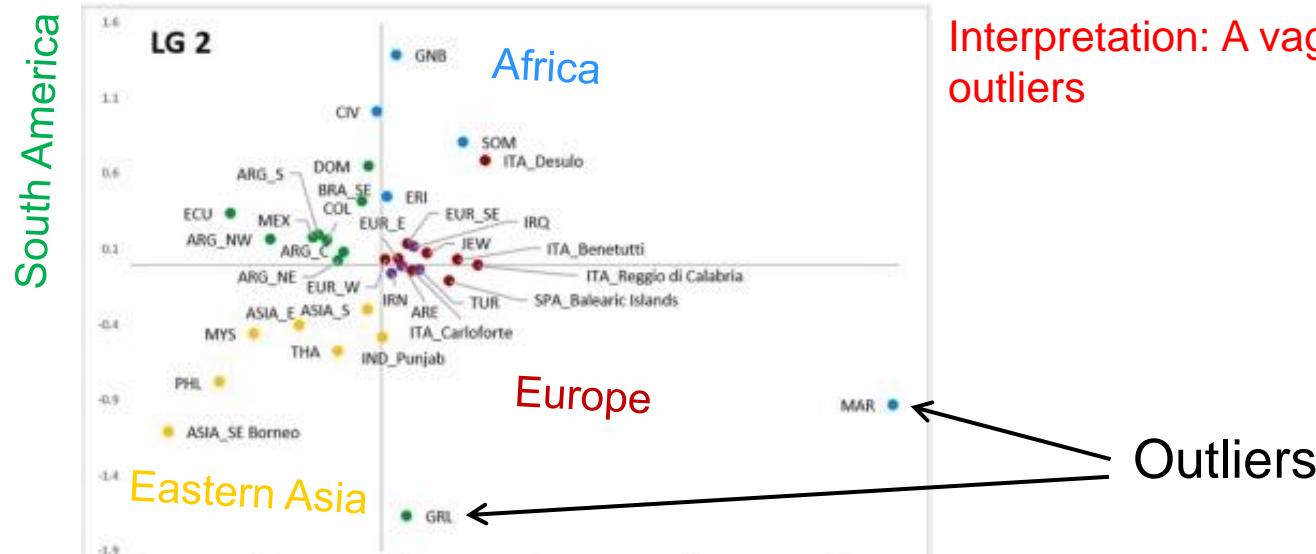
Show more 

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<https://doi.org/10.1016/j.fsigen.2025.103232> 

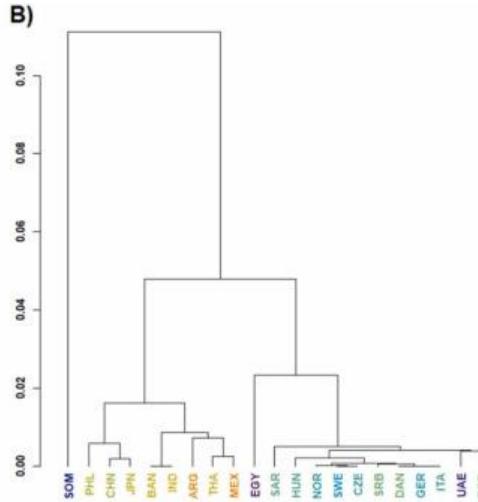
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# Differentiation



Interpretation: A vague separation with outliers

# Differentiation



Interpretation: Somali population is outlier. Asian populations cluster and European/Middle East

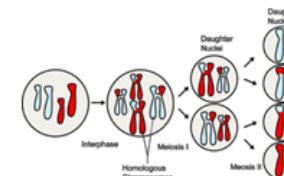
From Bergseth et al. 2023

# Linkage disequilibrium (LD)

Recall from first presentation

## Linkage and Linkage disequilibrium

- Linkage (or genetic linkage)
  - Can be described as the co-segregation of closely located loci within a family or pedigree.
  - **Effects the transmission probabilities!**
- Linkage disequilibrium (LD)
  - Allelic association.
  - Two alleles (at two different markers) which is observed more often/less often than can be expected.
  - **Effects the founder genotype probabilities, not the transmission probabilities!**
  - Haplotype frequencies rather than allele frequencies must be used



# Linkage disequilibrium (LD)

- Measures, e.g. D and r<sup>2</sup>

$$D = P_{A,B} - P_A P_B$$

$$r^2 = \frac{(P_{A,B} - P_A P_B)^2}{P_A P_B (1 - P_A) (1 - P_B)}$$

Example

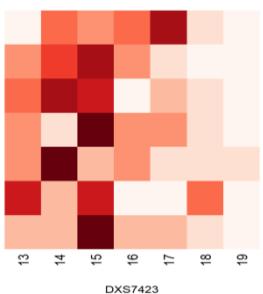
Haplotype	Observations	Ho	He
12 20	9	9/20	0.5*0.5
12 21	1	1/20	0.5*0.5
14 20	1	1/20	0.5*0.5
14 21	9	9/20	0.5*0.5

$$D=0.45-0.25=0.2$$

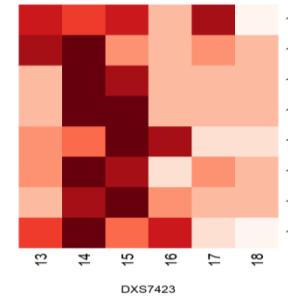
$$r^2=(0.45-0.25)^2/(0.5*0.5*0.5*0.5)=0.64$$

# Population genetics

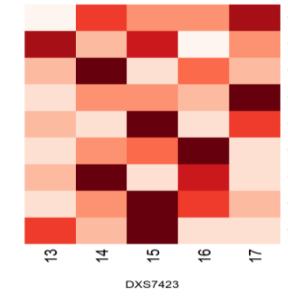
LD measured by D between pairs of alleles at two markers



Somalia



Norway



Sweden

Difficult to assess the overall LD

# Linkage disequilibrium (LD)

RESEARCH ARTICLES | JUNE 05 2003  
Volume 54, Issue 4  
May 2003



**Entropy as a Measure for Linkage Disequilibrium over Multilocus Haplotype Blocks**

Subject Area:  Genetics

M. Nothnagel; R. Fürst; K. Rohde

Hum Hered (2003) 54 (4): 186–198.  
<https://doi.org/10.1159/000070664> 

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## Abstract

**Objective:** The presence of linkage disequilibrium (LD) forms the basis for a range of uses, including the fine-mapping of diseases and studies on human genealogy. Recent findings indicate that single nucleotide polymorphisms (SNP) can occur in blocks of limited haplotypic diversity with high degrees of LD. Commonly used measures for LD, such as  $r^2$  and D', consider only two loci and might miss information to appropriately describe LD in larger haplotypic structures. **Methods:** We introduce the Normalized Entropy Difference, E, as a new multilocus measure for LD. A related quantity,  $\Delta S$ , provides an approximate  $\chi^2$  test for the significance of LD. The ability of the measure to detect LD in blocks is investigated using simulated data.

## Entropy

# Linkage disequilibrium (LD)

- Measures, e.g. D and r<sup>2</sup>

$$D = P_{A,B} - P_A P_B$$

$$r^2 = \frac{(P_{A,B} - P_A P_B)^2}{P_A P_B (1 - P_A) (1 - P_B)}$$

- What about multiallelic markers?

- We use entropy!

$$H = - \sum_s p_s \times \ln(p_s)$$

H is entropy and  $p_s$  is allele frequency

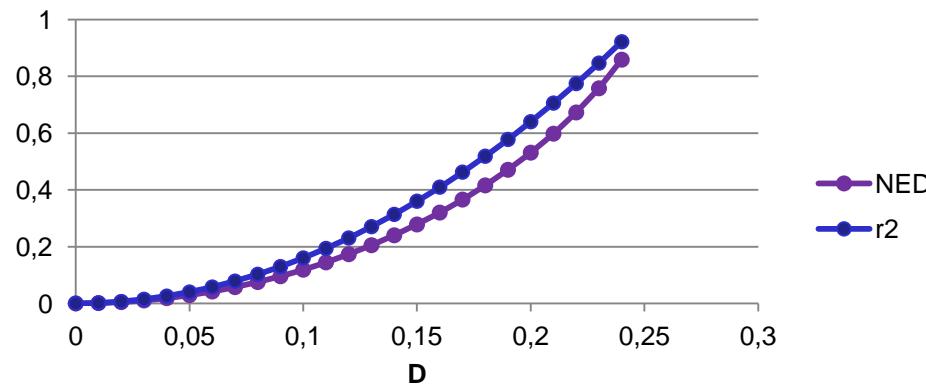
# Linkage disequilibrium

Normalized Entropy Difference (NED)

Single marker entropies

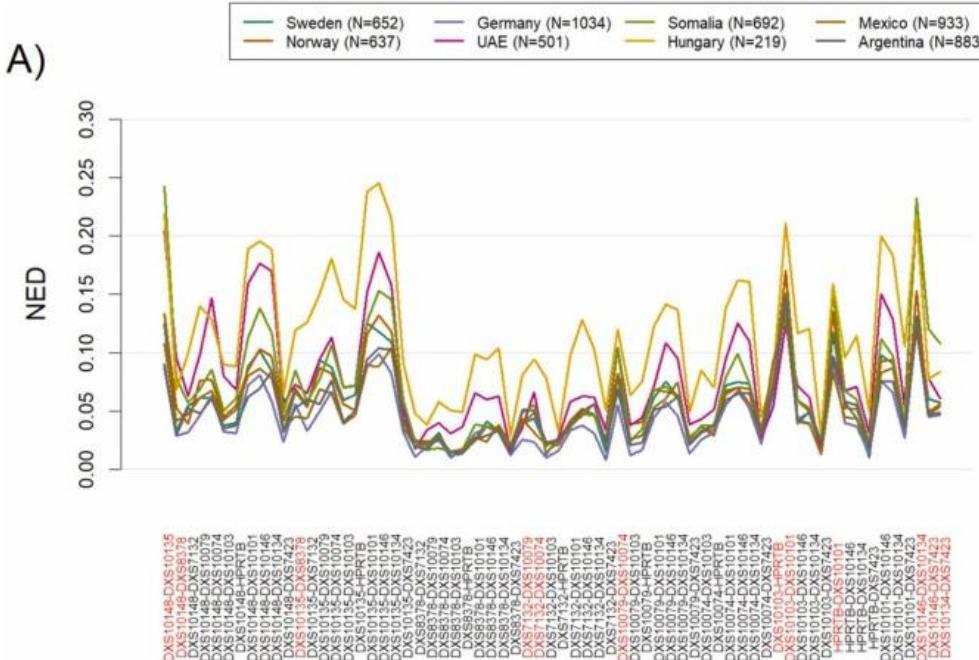
Joint entropy

$$NED_{i,j} = 2 \times \frac{(H_i + H_j) - H_{i,j}}{H_i + H_j}$$



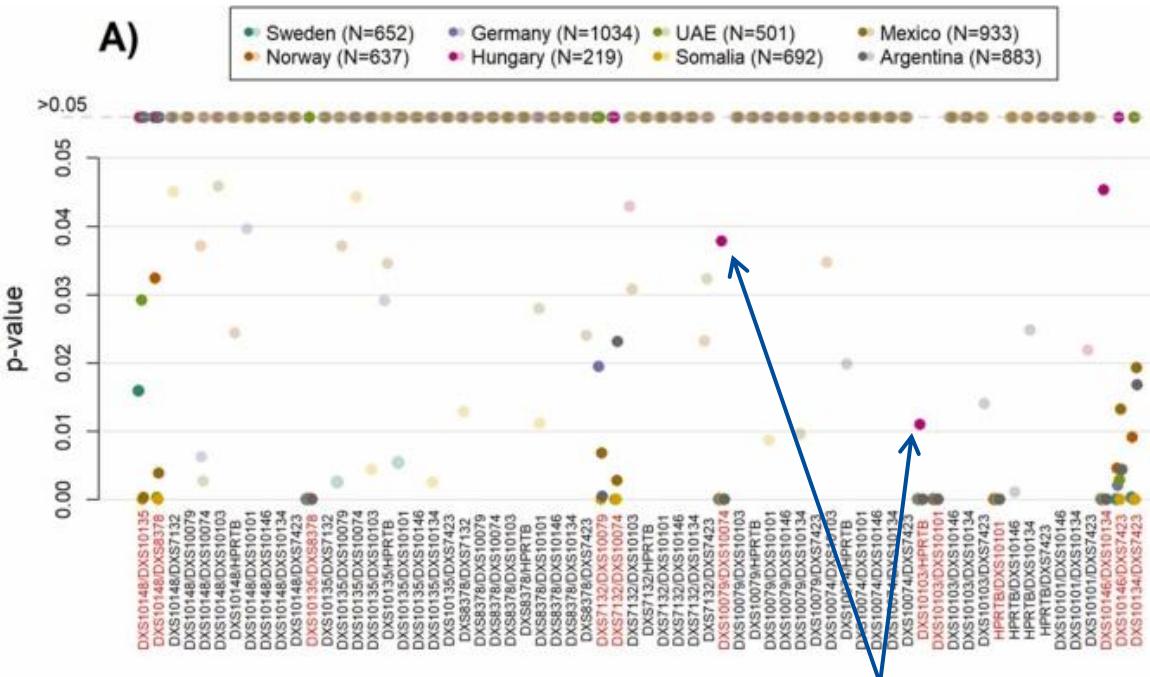
Two SNPs with minor allele frequencies of 0.5. NED as computed based on the degree of LD (D)

# Population genetics



Interpretation: Similar patterns of NED for the populations

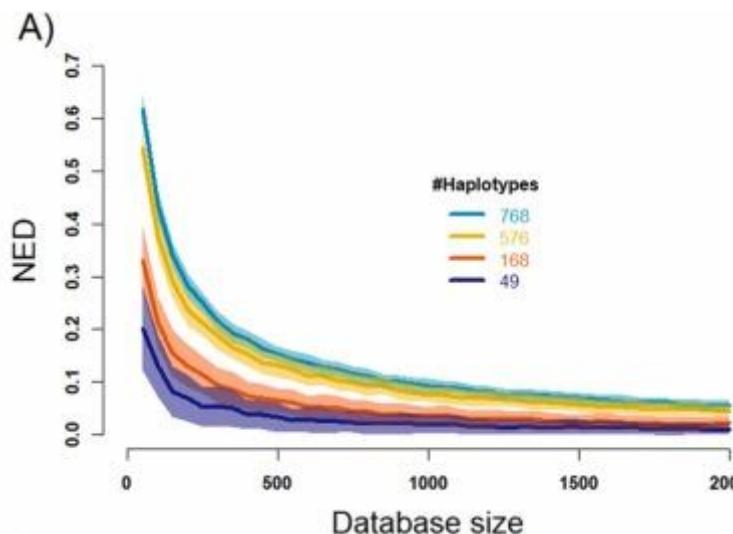
# Population genetics



Interpretation: Smaller databases may fail to detect the presence of LD

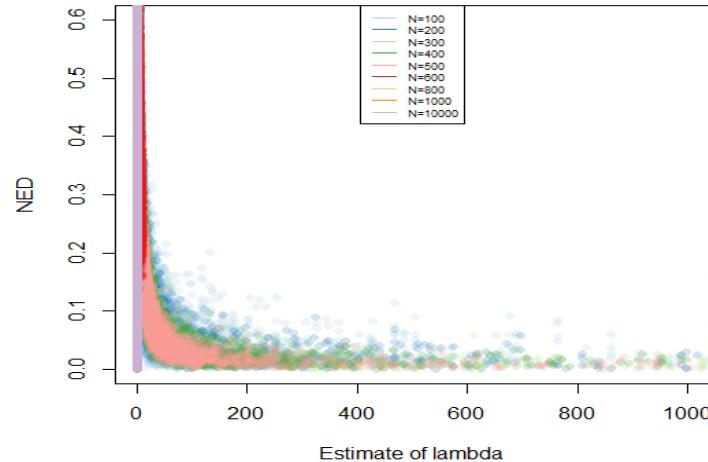
# Population genetics

Data based on linkage equilibrium (LE)



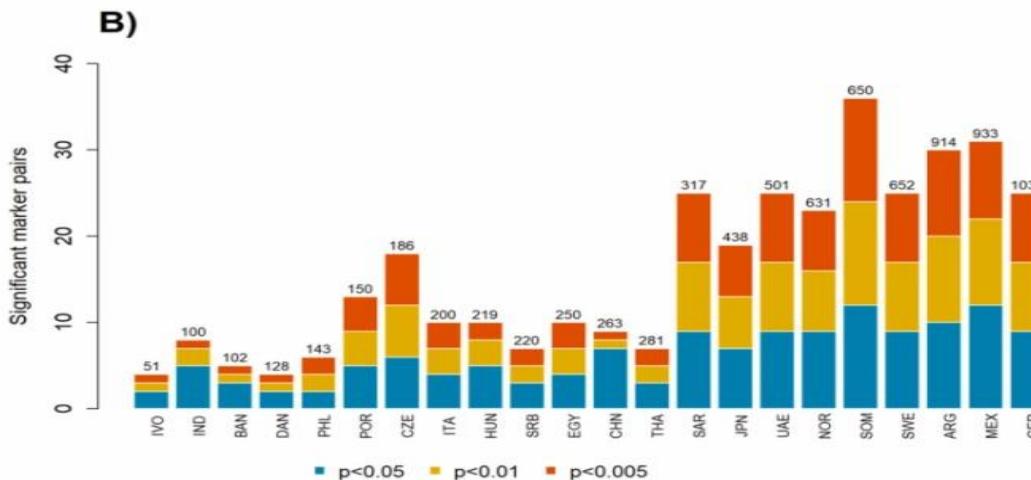
Interpretation: The smaller the sampled database, the higher the NED

# Population genetics



Interpretation: Large databases leads to low (zero) estimates of lambda

# On the importance of size



Forensic Science International: Genetics  
Volume 60, September 2022, 102745  
Research paper  
Extended population genetic analysis of 12 X-STRs – Exemplified using a Norwegian population sample  
Erik F. Bergseth<sup>a,1</sup>, Andreas Tillmar<sup>b,1</sup>, P. Jørgen T. Hadeland<sup>a</sup>, Daniel Kling<sup>a,b</sup>   
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<https://doi.org/10.1016/j.fsigen.2022.102745>   
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open access   
Highlights  
• Untraditional approaches to evaluating forensic markers.

From Bergseth et al. 2022

In small sized databases there is less LD??

# On the importance of size

## Example

Two SNPs with 2 alleles each => Total 4 possible haplotypes

Two STRs with 15 alleles each => Total of 225 possible haplotypes

# On the importance of size

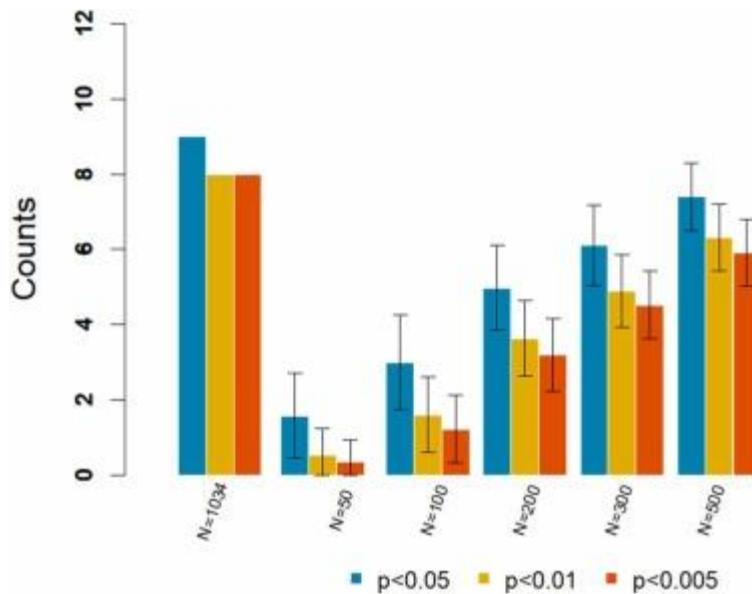
Table 2. Population genetic data for the Norwegian population sample (N = 631), with relevant forensic efficiency parameters for four triads of X-chromosomal STR markers in the Argus X-12 QS kit. MEC = Mean exclusion chance, PE=Power of Exclusion. PDF/PDM/MEC Desmarais Trio and Duo are calculated using formulas in Desmarais et al. [11] whereas PE (Paternal half sisters) and PE (Paternal grandmother/granddaughter) are calculated using formulas in Pinto et al. [12].

	Cluster 1	Cluster 2	Cluster 3	Cluster 4
Theoretical haplotypes	5376	1560	1197	3456
Observed haplotypes	362	204	193	275
Most common haplotype	7	20	24	13
Singlets	218	91	91	142
Power of Discrimination in Females (PDF)	0.999969	0.999791	0.99972	0.99991
Power of Discrimination in Males (PDM)	0.996074	0.989685	0.988032	0.993256
MEC Desmarais Trio	0.996059	0.989582	0.987896	0.993212
MEC Desmarais Duo	0.99217	0.979544	0.976307	0.986584
PE (Paternal half sisters)	0.984411	0.959637	0.953365	0.973399
PE (Paternal grandmother/granddaughter) <sup>a</sup>	0.992139	0.979344	0.976042	0.986496

5376 possible haplotypes!  
Only 362 observed

From Bergseth et al. 2022

# On the importance of size



From Bergseth et al. 2022

# Simulation as a mean to evaluate markers

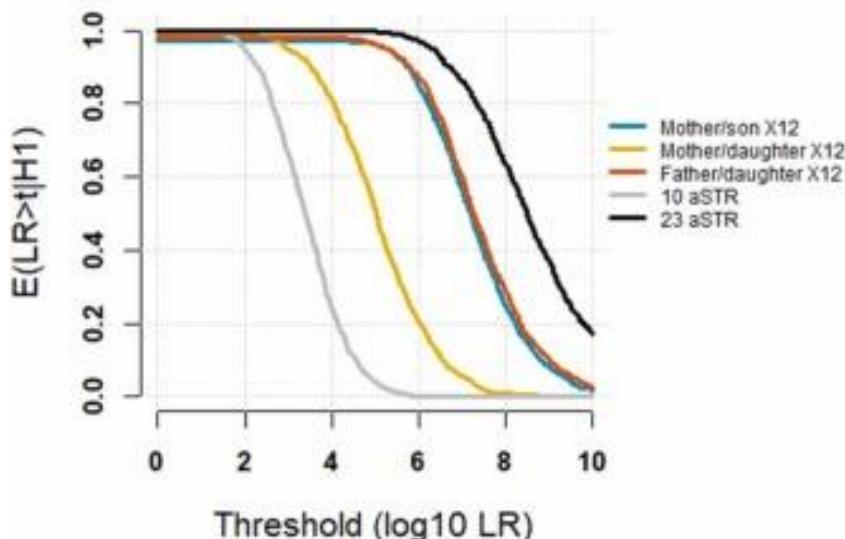
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Are these markers useful?

# Simulation as a mean to evaluate markers

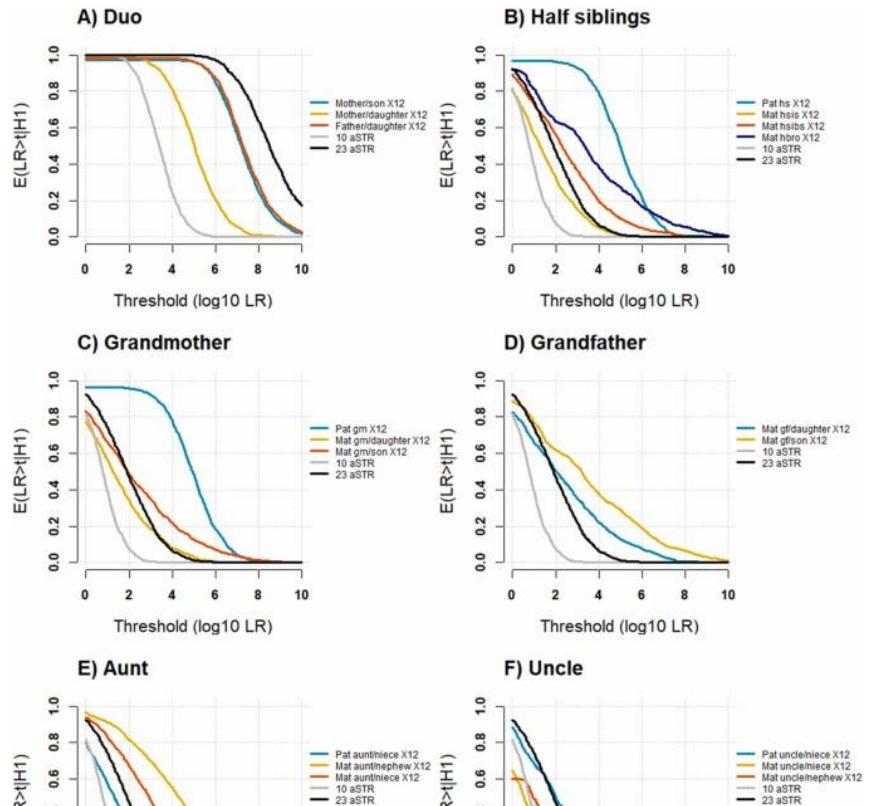
A) Duo



Conclusion: 12 X-STRs is more informative than 10 extra aSTRs

From Bergseth et al. 2022

# Simulation as a mean to evaluate markers



From Bergseth et al. 2022

# Haplotypes and databases

---

DANIEL KLING & ANDREAS TILLMAR

Daniel.kling@rmv.se

Andreas.tillmar@rmv.se



## ● WORKSHOP 1

X-chromosomal markers in forensic genetics  
Daniel Kling & Andreas Tillmar

## ● WORKSHOP 2

Acreditación en el campo de la Genética Forense y  
estrategias de validación de ensayos  
Manuel Crespillo Márquez, Rosalía Izquierdo & Estel Enricig Cabanes

## ● WORKSHOP 3

La genética en la Identificación de víctimas a gran escala:  
comparación de perfiles y evaluación estadística con Familias  
Carlos Vullo & Lourdes Prieto